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Model Study and Partial Synthesis of Prehispanolone and 14,15-Dihydroprehispanolone from Hispanolone!

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Abstract: Employing an intramolecular Michael addition as a pivotal step, furan 4 has been converted to dioxaspiro compounds 5 and 6, whose heterocyclic frameworks constitute important structural units of 14,15-dihydroprehispanolone 3 and prehispanolone 1, respectively. Hispanolone 2 was converted to 3 as well as 1 by utilizing a similar strategy.

We recently reported that hispanolone 2 was obtained through a mild acid hydrolysis of prehispanolone $1^{3,4}$ which was a specific platelet activating factor (PAF) receptor antagonist.^{5,6} Catalytic hydrogenation of 1 converted it to the bioactive and acid-insensitive 14,15-dihydroprehispanolone $3^{3,4,6}$. For further pharmacological evaluation purpose, it appears that both 1 and 3 are good leads for a structure-activity relationship study.

Scheme 1.

1, 0.5% HCl, EtOAc, (100%); ii, H₂, 5% Pd-C, EtOAc, (91%)

In view of its relative structural simplicity, furan 2 could serve as a key intermediate en route to the total synthesis of 1 and 3. For assessment reason, nevertheless, we also initiated a program to construct three model compounds, namely 5, 6 and 4. Here we report the synthesis of 5 and 6 from 4, as well as the construction of 1 and 3 from 2.

To our best knowledge, no arduous attempt to construct dioxaspiro[4,4]nonene framework7 similar to that of 1 has been recorded, notwithstanding that such spiro moiety is a common structural unit in many natural products.^{4,8} As outlined in Scheme 2, the synthesis of 4 from 3-furoic acid was straightforward.⁹ The deprotonation-silylation of furan 4,¹⁰ gave 11a and 11b in 1:1 ratio. Peracid oxidation¹¹ of a mixture of 11a

and 11b afforded the chromatographically separable 12a and 12b. Desilylaion of 12a furnished 13.¹² An intramolecular Michael addition of 13 gave 14.13 which was reduced 14.15 via 15 to give 5. On the other hand, phenyl sulfide 16 was prepared and subsequent oxidation and elimination¹⁶ eventually yielded 6.

Scheme 2.

i, LiAlH₄, Et₂O; ii, PPh₃, CBr₄, CH₂Cl₂, O°C; iii, NaH, PhCOCH₂CO₂Et, THF, O°C; iv, 5% NaOH, Ihen 2N HCl; v, PhLi, El₂O, -78°C; vi, 2.2 equiv n-BuLi,

Similar routes were utilized to construct 1 and 3 from the readily available $2³$. As shown in Scheme 3, protection of the keto group of 2 gave 17. Deprotonation and silylation of 17 yielded a mixture of 18a and 18b, which was not separated and was oxidized with peracid to furnish a chromatographically separable mixture of butenolides 19a and 19b. Desilylation of 19a converted it to the key intermediate 20, which underwent an intramolecular Michael addition to give again a chromatographically separable mixture of a pair of diastereomers 21a and 21b, the only stereochemical difference being the 13S or 13R configuration, respectively. It is significant to note that the biosynthetic pathway leading to the absolute configuration of the spiro carbon (C-13) in 1 is probably non-enantiospecific, because two related compounds, namely scutellone B and scutellone G have been identified.¹⁷

The $13S$ configuration of 21a was established by its DIBAL reduction, from which both 22 and a crystalline side product 23 were isolated. The X-ray crystallographic analysis of 23^{18} unequivocally certified its 135 configuration and further revealed that the molecular conformation is stabilized by intramolecular hydrogen bonding involving both hydroxyl groups, one of which acting as a donor in the bifurcated mode (Figure 1). Further conversion (deprotection and silane reduction) of 22 led to the "non-natural" $24,^{19}$ whose spiro carbon $C-13$ must be also of S configuration.

In principle, the verification of the 13S configuration of 21a also indirectly substantiated the

Figure 1. X-ray single crystal structure of 23.

13R configuration of 21b, which was likewise reduced to 25. Compound 3^{19} was obtained upon silane reduction and concomitant deprotection. The physical and spectroscopic properties of 3 were identical with those of a "natural" 3 obtained through catalytic hydrogenation of the natural 1.3 However, it was necessary to apply a modified Ley procedure¹⁶ to convert 25 to 1, presumbly due to the interference of the C-7 keto group regenerated during the conversion of 25 to 26. Thermal elimination of 26 eventually gave 1 , ¹⁹ whose physical and spectroscopic data were identical to those of the natural 1.³

Scheme 3.

i, (CH₂OH)₂, *p*-TsOH, C₈H₆, heat, ii, 2.2 equiv n-BuLi, C₈H₁₄, TMEDA, 0°C, then 2 5 equiv Me₃SiCi, Eb O, 0°C, iii, MeOO₃H, NaOAc, CH₂Cb,
7°C; iv, BF₃ Et O, CH₂Cl₂, -10°C, v, DBN-Et₃N: vi, -Bu₂ (EIO)₃P, PhMe, heat

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- 18. *Crystal data* for 23 (Siemens P4 system using Mo-K_{α} radiation. $\lambda = 0.71073$ A): C₂₂H₃₈O₅, M = 382.52. colorless orthorhombic prism, space group *P2,2,2, (No.* 19). *a =* 10.370(2), *b =* 11.785(2), c $= 17.454(3)$ Å, $\rho_{\text{calc}} = 1.191$ g cm⁻³, Z = 4, $F(000) = 840$, crystal size 0.20 x 0.42 x 0.60 mm. The structure was refined using SHELXTL-PLUS²⁰ for 1436 observed reflections $[2\theta_{\text{max}} = 50^{\circ}, |F_o| >$ $3\sigma(F_o|)$ and 242 variables to $R_F = 0.056$ and $R_{V} = 0.080$ with the weighting scheme $w = [\sigma^2(|F_o|) +$ 0.0013IF₀²]⁻¹ and an extinction parameter $\chi = 0.0016(5)$ where $F_c^* = F_c[1 + 0.002\chi F_c^2/\sin 2\theta]^{-1/4}$. Tables of atomic parameters have heen deposited at the Camhridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ. United Kingdom.
- 19. Selected physical and spectroscopic data : Compounds 1: oil; $[\alpha]_D^{25}$ -64.6^o (C₆H₆, c 0.85) [lit¹ $[\alpha]_D^{22}$ -63.6° (C₆H₆, c 0.55)]. Compound 3: oil: $\alpha \ln^{27}$ -32.2° (CHCl₃, c 0.72) [lit¹ $\alpha \ln^{22}$ -33.6° (CHCl₃, c 0.60)]. Compound 20: solid; m.p. 140-141°C; $[\alpha]_D^{25}$ 10.90° (CDCl₃; c 5.0). Compound 21a: oil; $[\alpha]_D^{24}$ 20.6° (CDCl₃; ϵ 4.35). Compound 21 b: oil; $[\alpha]_D^{24}$ -14.3° (CDCl₃; ϵ 4.60). Compound 24: oil; [a]\$' -Ill"(CHCl,. ~0.18); 'H NMR (CDCI,) 60.87 (s. 6H). 1.06 (d, *J6.5* Hz, 3H), 1.12 (s, 3H). 1.20-1.31 (m, 3H). 1.42-1.59 (m, 3H). 1.85-1.93 (m, 3H). 1.97-2.04 (m. 2H). 2.15-2.30 (m, 3H), 2.35-2.49 (m, 1H). 2.72 (y, *J6.5* Hz. IH), 3.51-3.68 (ABq, *J8.4* Hz, 2H), 3.79-3.90 (m, 2H); 13C NMR (CDCl₃) δ 8.74, 17.21, 18.13, 20.73, 29.10, 29.29, 32.12, 33.18, 37.63, 38.64, 39.76, 41.18, 42.36, 46.47, 50.01, 66.92, 77.32, 90.65, 95.94, 210.20.
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